



Amphetamine Direct ELISA Kit

Catalog No. BQ 209-096 (96 tests)

(hydroxy methoxyamphetamine) Phenteramine	28	89
Fenfluramine	>2500	<1
d-Ephedrine	>2500	<1
l-Ephedrine	>2500	<1
d-Phenylpropanolamine	>2500	<1
l-Phenylpropanolamine	>2500	<1
dl-MDMA	>2500	<1
(methylene dioxymethamphetamine) dl-MDEA	>2500	<1
(methylene dioxyethylamphetamine) d-pseudoephedrine	>2500	<1
l-Pseudoephedrine	>2500	<1
dl-MBDB	>2500	<1
(N-methyl-3,4methylene dioxyphenyl-2-butamine)Tyramine	>2500	<1
Methylphenidate	>2500	<1

6. Cross-Reactivity with Unrelated Drugs

Aliquots of a human urine matrix were spiked with the following compounds at a concentration of 5,000 ng/ml. None of these compounds gave values in the assay that were equal to or greater than the assay sensitivity level (1 ng/ml).

Acetaminophen, Acetylsalicylic acid, Aminopyrine, Ampicillin, Amobarbital, Ascorbic acid, Atropine, Barbitol, Benzoylcegonine, Butabarbital, Caffeine, Cocaine, Carbamazepine, Codeine, Chloroquine, Chlorpromazine, Carbromal, Desipramine, Dextromethorphan, Dextropropoxyphene, 5,5-Diphenylhydantoin, 10-11-Dihydrocarbamazepine, Diazepam, Ethosuximide, Estriol, Estrone, Estradiol, Ethotoin, Glutethimide, Hexobarbital, Ibuprofen, Imipramine, Lidocaine, LSD, Methadone, Methadone-primary metabolite, Methaqualone, Methbarbital, Mephentoin, "Methyl"-propylsuccinimide, Mephobarbital, Methyl PEMA, Methsuximide, 4-Methylprimidone, Morphine, Meperidine, Niacinamide, Norethindrone, N-Normethsuximide, Phenobarbital, Phensuximide, PEMA, Primidone, Phencyclidine, Pentobarbital, Phenothiazine, Phenylpropanolamine, Procaine, Quinine, Secobarbital, Tetracycline, Tetrahydrozoline, THCCOOH

REFERENCES

1. Urine Testing for Drugs of Abuse, National Institute on Drug Abuse Research Monograph. 73: 95-97 (1986).
2. N.Weiner. Norepinephrine, epinephrine and the sympathomimetic amines. In: The Pharmacological Basis of Therapeutics. 7th ed. p.145-180 (New York: MacMillan 1985).
3. J. Caldwell and P.S. Sever. The Biochemical Pharmacology of Abused Drugs. Clinical Pharmacology and Therapeutics. 16: 625- 638 (1974).
4. R. C. Baselt. In: Advances in Analytical Toxicology, Vol.1. p.87 - 93. Ed. R. C. Baselt, Biomedical Publications, Foster City, CA (1984).

2010-01-25 (Mfg:06/2001)

Warning

All of Bio-Quant ELISA Kits have not been tested for clinical use and are not approved in the United States by the FDA for diagnostic clinical use. They are components or reagents made solely for research use, further manufacturing and export use. It is the commitment of Bio-Quant customers to receive its products solely for the purpose of exportation or research, and not for the purposes of clinical diagnostic use.

BIO-QUANT DOES NOT MAKE ANY OTHER WARRANTY OR REPRESENTATION WHATSOEVER, WHETHER EXPRESS OR IMPLIED, WITH RESPECT TO THESE PRODUCTS. IN PARTICULAR Bio-Quant, INC. DOES NOT MAKE ANY WARRANTY OF SUITABILITY, NON-INFRINGEMENT, MERCHANTABILITY OR FITNESS FOR ANY PARTICULAR PURPOSE OF ANY PRODUCT

INTENDED USE

The Amphetamine Direct ELISA Kit provides only a preliminary analytical test result. A more specific alternate chemical method must be used in order to obtain a confirmed analytical result. Gas chromatography/ mass spectrometry (GS-MS) is the preferred confirmatory method (1). Professional judgment should be applied to any drug of abuse test result, particularly when preliminary positive results are used.

SUMMARY AND EXPLANATION

The Amphetamine Direct ELISA Kit is a specific and sensitive in-vitro test to detect the presence of d-amphetamine in samples such as whole blood, oral fluids, serum, plasma and urine. While the assay will detect amphetamine use, interference by l-amphetamine and pseudo-ephedrine is virtually nonexistent. Amphetamine is a potent central nervous system stimulant. The (+)-isomer also referred to as d-amphetamine is three to four times more potent than the (-)-isomer, l-amphetamine (2). Amphetamine may be metabolized and excreted as the p-hydroxy isomer. Amphetamines act by inducing euphoria, irritability, anxiety and paranoia. Urinary excretion rates are influenced by the urinary pH with acidic urine favoring the excretion of unchanged drug(2). Up to 80% of a given dose may be excreted unchanged, especially in acid urine. Alkaline urine reduces the excretion of unchanged amphetamine to less than 5% of the dose.

PRINCIPLES OF THE TEST

The Amphetamine Direct ELISA Kit (for d-amphetamine measurement) is based upon the competitive binding to antibody of enzyme labeled antigen and unlabeled antigen, in proportion to their concentration in the reaction mixture. A 10 µl. aliquot of a diluted unknown specimen is incubated with a 100 µl. dilution of enzyme (Horseradish peroxidase) labeled d-amphetamine derivative in micro-plate wells, coated with fixed amounts of oriented high affinity purified polyclonal antibody. The wells are washed thoroughly and a chromogenic substrate added. The color produced is stopped using a dilute acid stop solution and the wells read at 450 nm. The intensity of the color developed is inversely proportional to the concentration of drug in the sample. The technique is sensitive to 1 ng/ml. The Amphetamine Direct ELISA Kit avoids extraction of urine sample for measurement. It employs a d-amphetamine directed antiserum. Due to the proprietary method of orienting the antibody on the polystyrene micro-plate much higher sensitivity is achieved compared to passive adsorption. This allows an extremely small sample size reducing matrix effects and interference with binding proteins(s) or other macromolecules.

MATERIALS PROVIDED	96Test Kit
1. microwells coated with polyclonal anti-d-amphetamine	12x8x1
2. d-Amph-Conjugate	12.5 ml
3. Immunalysis Positive Reference Standard	1ml
4. Negative Standard	1 ml
5. TMB Substrate	14 ml
6. Stop Reagent	12.5 ml

MATERIALS NOT PROVIDED

1. Distilled or deionized water
2. Precision pipettes
3. Disposable pipette tips
4. ELISA reader capable of reading absorbance at 450 nm
5. Absorbance paper or paper towel
6. Graph paper

STORAGE AND STABILITY

1. The expiration date of the kit is stated on the label. The kit can be expected to perform satisfactory until the expiration date is stored in the refrigerator at 2-40° C.
2. Store the kit at 2-8° C.
3. Keep microwells sealed in a dry bag with desiccants.
4. The reagents are stable until expiration of the kit.
5. Do not expose test reagents to heat, sun or strong light.
- 6.

WARNINGS AND PRECAUTIONS

1. Potential biohazardous materials:
The calibrator and controls contain human source components which have been tested and found non-reactive for hepatitis B surface antigen as well as HIV antibody with FDA licensed reagents. However, as there is no test method that can offer complete assurance that HIV, Hepatitis B virus or other infectious agents are absent, these reagents should be handled at the Biosafety Level 2, as recommended in the Centers for Disease Control/National Institutes of Health manual, "Biosafety in Microbiological and Biomedical Laboratories." 1984
2. This test kit is designed for research use only.
3. Do not pipette by mouth. Do not smoke, eat, or drink in the areas in which specimens or kit reagents are handled.
4. The components in this kit are intended for use as an integral unit. The components of different lots should not be mixed.
5. It is recommended that serum samples be run in duplicate.
6. Optimal results will be obtained by strict adherence to this protocol. Accurate and precise pipetting, as well as following the exact time and temperature requirements prescribed are essential. Any deviation from this may yield invalid data.

SPECIMEN COLLECTION HANDLING

1. The Amphetamine Direct ELISA Kit is to be used with human samples, such as whole blood, oral fluids, serum, urine and plasma. has not tested all possible applications of this assay. Cutoff criteria are important in deciding the sample dilution.
2. Specimens to which sodium azide has been added affect the assay.
3. Urine samples should be stored at 2 - 40 C until use. Samples should be well mixed before assay.
4. Repeated freezing and thawing should be avoided. Urine samples should be shipped refrigerated with Blue Ice or equivalent.

ASSAY PROCEDURE

All reagents must be brought to room temperature (18-26° C) before use. The procedure as described below may be followed in sequence using manual pipettes. Alternatively all reagents may be added using an automated pipettor.

- 1) Dilute specimens, to the necessary range with Phosphate Buffered Saline pH 7.0. (Urine samples 1:20 for a cutoff level of 500ng/ml.) The dilution factor can be adjusted based on the laboratories cutoff.
- 2) Add 10ul of appropriately diluted calibrators and standards to each well in duplicate.
- 3) Add 10ul of the diluted specimens in duplicate (recommended) to each well.
- 4) Add 100ul of the Enzyme Conjugate to each well. Tap the sides of the plate holder to ensure proper mixing.
- 5) Incubate for 60 minutes at room temperature preferably in the dark (18-26° C), after addition of enzyme conjugate to the last well.
- 6) Wash well 6 times with 350ul distilled water using either a suitable plate washer or wash bottle taking care not to cross contaminate wells. If testing samples containing abnormally high amount of hemoglobin (some postmortem samples) use 10mM Phosphate buffered saline pH 7.0-7.4. This will lower potential nonspecific binding of hemoglobin to the well, thus lowering background color.
- 7) Invert wells and vigorously slap dry on absorbent paper to ensure all residual moisture is removed. This step is critical to ensure that residual enzyme conjugate, does not skew results. If using an automated system, ensure that the final aspiration on the wash cycle aspirates from either side of the well.
- 8) Add 100ul of Substrate reagent to each well and tap sides of plate holder to ensure proper mixing.
- 9) Incubate for 30 minutes at room temperature, preferably in the dark.
- 10) Add 100ul of Stop Solution to each well, to change the blue color to yellow.
- 11) Measure the absorbance at a dual wavelength of 450 nm. and 650 nm.

- 12) Wells should be read within 1 hours of yellow color development.

The following data represent a typical dose/response curve.

d-amphetamine (ng/ml)	Absorbance
0	2.459
10	0.891
25	0.431
50	0.255

The dose/response curve shown above should not be used in assay calculations. It is recommended that at least one in-house positive quality control sample be included with every assay run. A dose response curve or a cutoff calibrator should be run with every plate.

RESULTS

If the average sample absorbance is equal to or less than the average absorbance of the laboratory positive reference standard the sample is POSITIVE for amphetamine. If the average sample absorbance is greater than the average absorbance of the laboratory positive reference standard the sample is called NEGATIVE for amphetamine. Alternatively a dose response curve can be established by plotting standard concentration (abscissa) against corresponding absorbance (ordinate). Values for unknown samples are obtained by interpolation from the curve

SPECIFIC PERFORMANCE CHARACTERISTICS

1. **Accuracy**
Forty whole blood samples and 40 urine samples collected from presumed non-users were tested in the Amphetamine Direct ELISA Kit. One hundred percent of these normal samples measured negative at 50 ng/ml for whole blood and 500 ng/ml for urine. Thirty five whole blood samples which were previously confirmed positive for amphetamine by GC-MS employing a cut-off of 50 ng/ml, were tested in the Amphetamine Direct ELISA Kit. All of the samples were found to be positive i.e. above the cut-off of 50 ng/ml.
2. **Precision**
The precision of the Amphetamine Direct ELISA Kit has been verified by assessment of the mean, standard deviation (SD) and coefficients of variation (CV) in data resulting from repetitive assays.
3. **Intra-assay Precision**
Intra-assay precision was determined with reference controls. A 0, 10, 25 and 50 ng/ml standard was assayed five times in the same assay. The results are tabulated in

Amphetamine (ng/ml)	Mean Abs.	S.D.	C.V.%
0	2.399	0.115	4.8
10	0.897	0.095	10.6
25	0.458	0.061	13.32
50	0.271	0.022	8.12

4. **Sensitivity**
Assay sensitivity based on the minimum amphetamine concentration required to produce a four standard deviation from assay Ao is 1 ng/ml.
5. **Specificity**
The specificity of the ELISA for Amphetamine was determined by generating inhibition curves for each of the compounds listed below the antisera cross-reactivities are listed in Table 2.

Compound	Approx. ng/ml equivalent to 25ng amphetamine	Cross-reactivities percentage
l-Amphetamine	865	2.9
HydroxyamphetamineHCl	57	44
l-Methamphetamine HCl	1250	2
dl-MDA	10	250
(methylenedioxyamphetamine) d-Methamphetamine HCl	417	6.5
dl- HMA	100	25