



6330 Nancy Ridge Drive Suite 103
San Diego, CA 92121
(858) 450-0048

Alkaline Phosphatase Reagent Set (Kinetic Procedure)

Catalog Number: BQ003C-CR

INTENDED USE

For the quantitative determination of alkaline phosphatase in human serum.

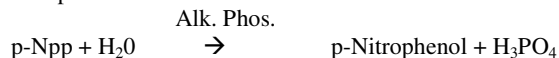
INTRODUCTION

Distributed in almost every tissue of the body, serum alkaline phosphatase (ALP) levels are of interest in the diagnosis of hepatobiliary disorder and bone disease.¹ Most of the ALP in the normal adult serum is from the liver or biliary tract.² Normal alkaline phosphatase levels are age dependent, and levels are elevated during periods of active bone growth. Moderate elevations of ALP (not involving the liver or bone) may be attributed to Hodgkin's disease, congestive heart failure, and abdominal bacterial infections.³ Elevations also occur in the third trimester of pregnancy.

Alkaline phosphatase is determined by measuring the rate of hydrolysis of various phosphate esters. p-Nitrophenyl Phosphate is one such ester that was used as a substrate by Fujita in 1939.⁴ Bowers and McComb further modified the procedure to a kinetic assay.⁵ In 1974, the Committee on enzymes of the Scandinavian Society for Clinical Chemistry and Clinical Physiology adopted a modification of the above procedure as the recommended procedure.⁶ The present method is a modification of the reference methods of the above committee and the American Association for Clinical Chemistry.⁷

PRINCIPLE

The enzymatic sequence employed in the assay of Alkaline Phosphatase is as follows:



p-Npp is colorless but p-Nitrophenol has strong absorbance at 405 nm. The rate of increased absorbance at 405 nm is proportional to the enzyme activity.

REAGENT COMPOSITION

When reconstituted as directed, the reagent for Alkaline Phosphatase contains the following:

(Alkaline Phosphatase Reagent): p-Nitrophenyl Phosphate 17mM, Magnesium Ions 4mM, Buffer (pH 10.2 ± 0.2), activator and binder.

WARNINGS AND PRECAUTIONS

1. For in vitro diagnostic use. CAUTION: In vitro diagnostic reagents may be hazardous. Handle in accordance with good laboratory procedures which dictate avoiding ingestion, and eye or skin contact.
2. Specimens should be considered infectious and handled appropriately.
3. Use distilled or deionized water indicated.

STORAGE AND STABILITY

Store reagent set at 2 – 8 °C (refrigerated). Reconstituted reagent is stable for thirty (30) days when stored at 2 – 8 °C and twenty-four (24) hours at room temperature.

REAGENT DETERIORATION

The reagent should be discarded if:

1. Turbidity has occurred; turbidity may be a sign of contamination.
2. Moisture has penetrated the vial and caking has occurred.
3. The reconstituted reagent has an absorbance against water greater than 0.8 at 405nm.

SPECIMEN COLLECTION

Unhemolyzed serum is the preferred sample. Heparinized plasma may also be used. Oxalate, fluoride and EDTA inhibit alkaline phosphatase, so are unsuitable as anticoagulants.⁸ Samples should be kept cold and assayed as soon as possible after collection. A timed routine for sample collection and analysis should be established in each laboratory because ALP levels in serum or plasma, or in reconstituted control serum, rise significantly when stored at 2 – 8 °C or at room temperature.

INTERFERING SUBSTANCES

EDTA, citrate, fluoride and oxalate inhibit alkaline phosphatase. Young et al. give a list of drugs and other substances, which may interfere with the determination of ALP activity.⁹

MATERIALS REQUIRED BUT NOT PROVIDED

1. Pipetting devices.
2. Test tubes/rack.
3. Timer.
4. Spectrophotometer with a temperature controlled cuvette.
5. Heating bath/block.

GENERAL INSTRUCTIONS

The reagent for Alkaline Phosphatase is intended for use either as an automated procedure on chemistry instruments or as a manual procedure on a suitable spectrophotometer.

PROCEDURE (AUTOMATED)

Refer to appropriate application manual available.

PROCEDURE (MANUAL)

1. Reconstitute reagent according to instructions.
2. Pipette 1.0 ml of reagent into appropriate tubes and allow to equilibrate to 37 °C.
3. Zero spectrophotometer with water at 405 nm.
4. Transfer 0.025 ml (25 µl) of sample to reagent. Mix well.
5. After one (1) minute, measure the absorbance. Return tube to 37 °C. Repeat readings every minute for the next two (2) min.*
6. Calculate the average absorbance difference per minute
7. The $\Delta\text{Abs.} / \text{min.}$ multiplied by the factor 2187 (see Calculations) will yield results in IU/L.
8. Samples with values above 800 IU/L should be diluted 1:1 with saline, re-assayed and the results multiplied by two (2).

Note: If the spectrophotometer being used is equipped with a temperature controlled cuvette. The reaction mixture may be left in the cuvette while the absorbance readings are taken.

VOLUMES

If the spectrophotometer being used can accurately read the final volume of 0.50ml or smaller, follow the "ALTERNATE PROCEDURE."

Unit: One unit is the amount of enzyme that catalyzes the transformation of one micromole of substrate per minute under the specified conditions.

$$\text{IU/L} = \frac{\Delta \text{Abs./min.} \times 1000 \times \text{TV}}{\epsilon \times \text{LP} \times \text{SV}} = \frac{\Delta \text{Abs./min.} \times 1000 \times 1.025}{18.75 \times 1 \times 0.025}$$

$$= \Delta \text{Abs./min.} \times 2187$$

Where:

$\Delta \text{Abs./min.}$ = Absorbance change per minute

1000 = Conversion of IU/ml to IU/L

TV = Total reaction volume (1.025 ml)

ϵ = Millimolar absorptivity of p-Nitrophenol (18.75)

LP = Light path in centimeters (1.0 cm)

SV = Sample volume (0.01 ml)

Example: If the $\Delta \text{Abs./min.} = 0.007$ then $0.007 \times 2187 = 15.3 \text{ IU/L}$.

NOTE: If test parameters are altered the factor has to be recalculated using the above formula

SI UNITS: To convert to SI Units (nkat/L) multiply IU/L by 16.67.

ALTERNATE PROCEDURE

1. Reconstitute reagent according to instructions.
2. Pipette 0.50ml (500 μL) of reagent into appropriate tubes and allow to equilibrate to 37 °C.
3. Zero spectrophotometer with water at 405nm.
4. Add 0.010 ml (10 μL) of sample to reagent, mix well.
5. After one (1) minute, measure the absorbance. Return tube to 37 °C. Repeat readings every minute for the next two (2) min.
6. Calculate the average absorbance difference per minute ($\Delta \text{Abs./min.}$).
7. The $\Delta \text{Abs./min.}$ multiplied by the factor 2720 (see Calculations) will yield results in IU/L.
8. Samples with values above 800 IU/L should be diluted 1:1 with saline, re-assayed and the results multiplied by two (2).

PROCEDURE CALCULATION

Unit: One unit is the amount of enzyme that catalyzes the transformation of one micromole of substrate per minute under the specified conditions.

$$\frac{\Delta \text{Abs./min.} \times 1000 \times 0.510}{\Delta \text{Abs./min.} \times 2720} = 1 \times 18.75 \times 1 \times 0.010$$

Where:

$\Delta \text{Abs./min.}$ = Absorbance

Change per minute. 1000 = Conversion of IU/ml to IU/L

TV = Total reaction volume (0.510 ml)

1 cm = Light path in centimeters

18.75 = Millimolar absorptivity of p-Nitrophenol

SV = Sample volume (0.025 ml)

Example:

$$A_1 = 0.56, A_2 = 0.60 \text{ then: } (0.60 - 0.56) = 0.04 \times 2720 = 108.8 \text{ IU/L}$$

NOTE: If test parameters are altered the factor has to be recalculated using the above formula

SI UNITS: To convert to SI Units (nkat/L) multiply IU/L by 16.67.

PROCEDURAL LIMITATIONS

This methodology measures total Alkaline Phosphatase irrespective of tissue or organ of origin. Further tests may be necessary to assist in differential diagnosis.

QUALITY CONTROL

It is recommended that controls be included in each set of assays. Commercially available control material with established Alkaline Phosphatase values may be used for quality control. The assigned value of the control material must be con/L, by the chosen application. Failure to obtain the proper range of values in the assay of control material may indicate either reagent deterioration, instrument malfunction, or procedural errors.

EXPECTED VALUES

Adults 25 - 90 IU/L at 37 °C. Children have a higher normal value. It is strongly suggested that each laboratory establish its own normal range.¹⁰

PERFORMANCE CHARACTERISTICS

1. Linearity: 900 IU/L
2. Comparison: A study performed between the present procedure and one commercial product resulted in a coefficient of correlation of 0.99 with a regression of $y = 0.95x + 3.50$.
3. Precision studies:

<i>Within Run</i>		
<u>Mean IU/L</u>	<u>S.D.</u>	<u>C. V.</u>
76.1	2.1	2.7%
331.3	15.4	4.6%

<i>Run to Run</i>		
<u>Mean IU/L</u>	<u>S.D.</u>	<u>C. V.</u>
75.3	5.1	6.7%
328.6	11.26	3.4%

REFERENCES

1. Kochmar, J.F., and Moss, D.W.: Fundamentals of Clinical Chemistry, N.W. Tietz (ed), p. 604, W.B. Saunders and Company, Philadelphia, PA (1976).
2. Kaplan, M.M., and Righetti, A.: J. Clin. Inv. 34:126 (1955).
3. Kaplan, M.M.: New England J. Med. 286:200 (1972).
4. Fugita, H.: Uber die Miorobestimmung der Blutphosphatase. J. Biochem. (Japan) 30:69 (1939)
5. Bowers, G.N., Jr., McComb, R.B.: A Continuous Spectrophotometric Method for Measuring the Activity of Serum Alkaline Phosphatase. Clin. Chem. 12:70 (1966).
6. The Committee On Enzymes of The Scandinavian Society for Clinical Chemistry and Clinical Physiology: Recommended Methods for the Determination of Four enzymes in blood. Scand. J. Clin. Lab. Invest. 32:29 (1974).
7. Tietz, N.W., (ed): Study Group on Alkaline Phosphatase. A Reference Method for measurement of alkaline phosphatase activity in Human Serum. Clin. Chem. 29:751 (1983).
8. Demetrious, J.A., et. al.: Enzymes in Clinical Chemistry Principles and Technics, 2nd Ed. Hagerstown (MD), Harper and Row 927 (1974).
9. Young, D.S., et al: Clin. Chem 21:5 (1975).
10. Tietz, N.: Fundamentals of Clinical Chemistry 602-609 (1976).